

CRISPR/Cas9 editing to produce low asparagine, low acrylamide wheat: out of the lab and into the field

Nigel G. Halford
Rothamsted Research

9th Plant Genomics & Gene Editing Congress Europe
April 11th – 12th, 2022



ROTHAMSTED
RESEARCH



Biotechnology and
Biological Sciences
Research Council

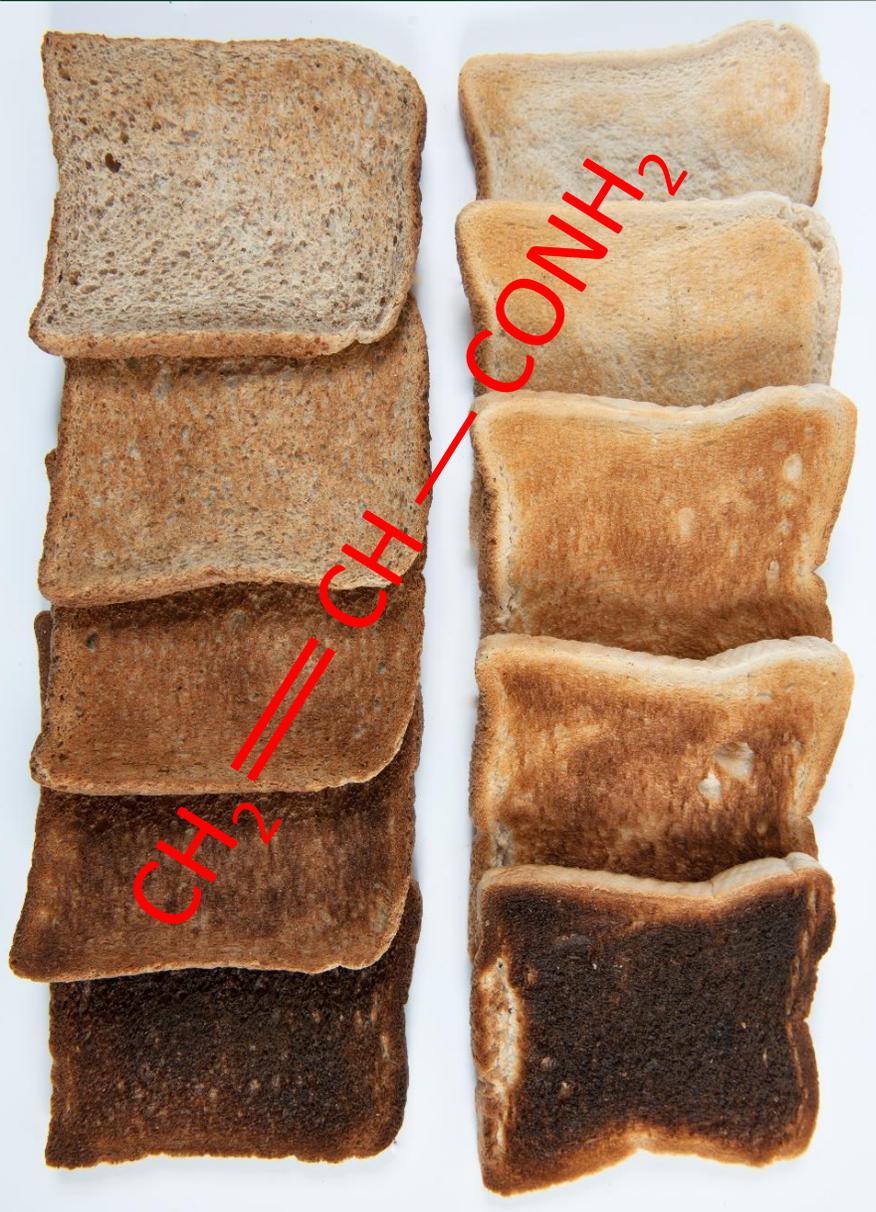


Lawes
Agricultural
Trust

Acrylamide



ROTHAMSTED
RESEARCH



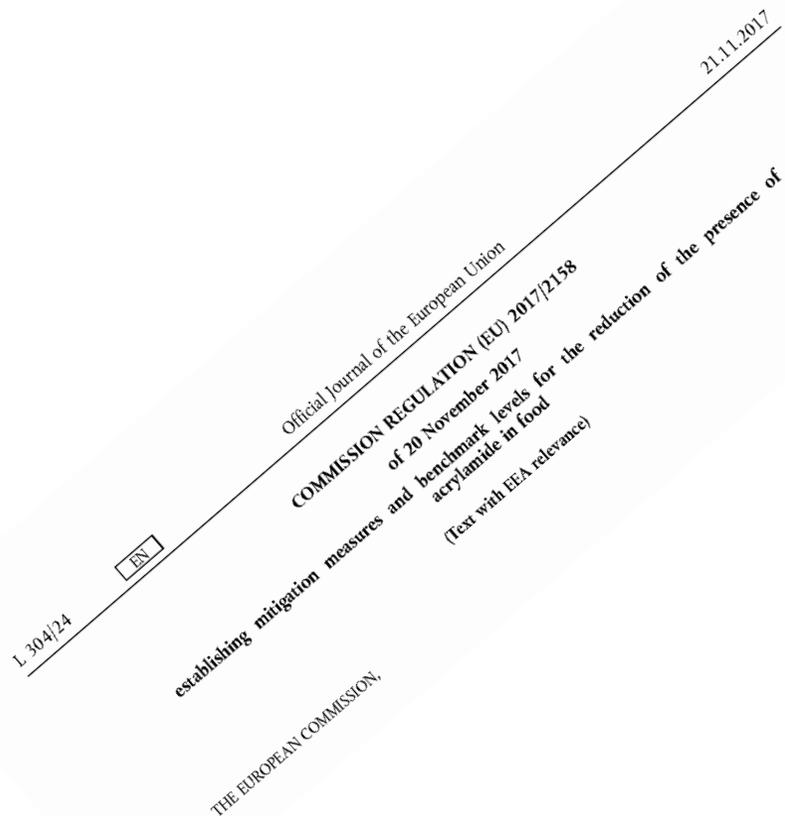
- The processing contaminant, acrylamide, was unexpectedly found in cooked foods, mainly those derived from plants, in 2002.
- Acrylamide causes cancer in rodents and is regarded as a probable (Group 2a) carcinogen in humans. It also affects development, and at high doses the nervous system and fertility.
- Fried, baked and roasted potato and cereal products and coffee are the major contributors to dietary intake in Europe.
- Bread (especially toasted), breakfast cereals, biscuits, cereal snack products, cakes, pies, batter, as well as chips (French fries), crisps, roast potatoes, and all types of coffee are all affected.



Commission Regulation (EU) 2017/2158: New risk management measures for acrylamide in Europe



ROTHAMSTED
RESEARCH

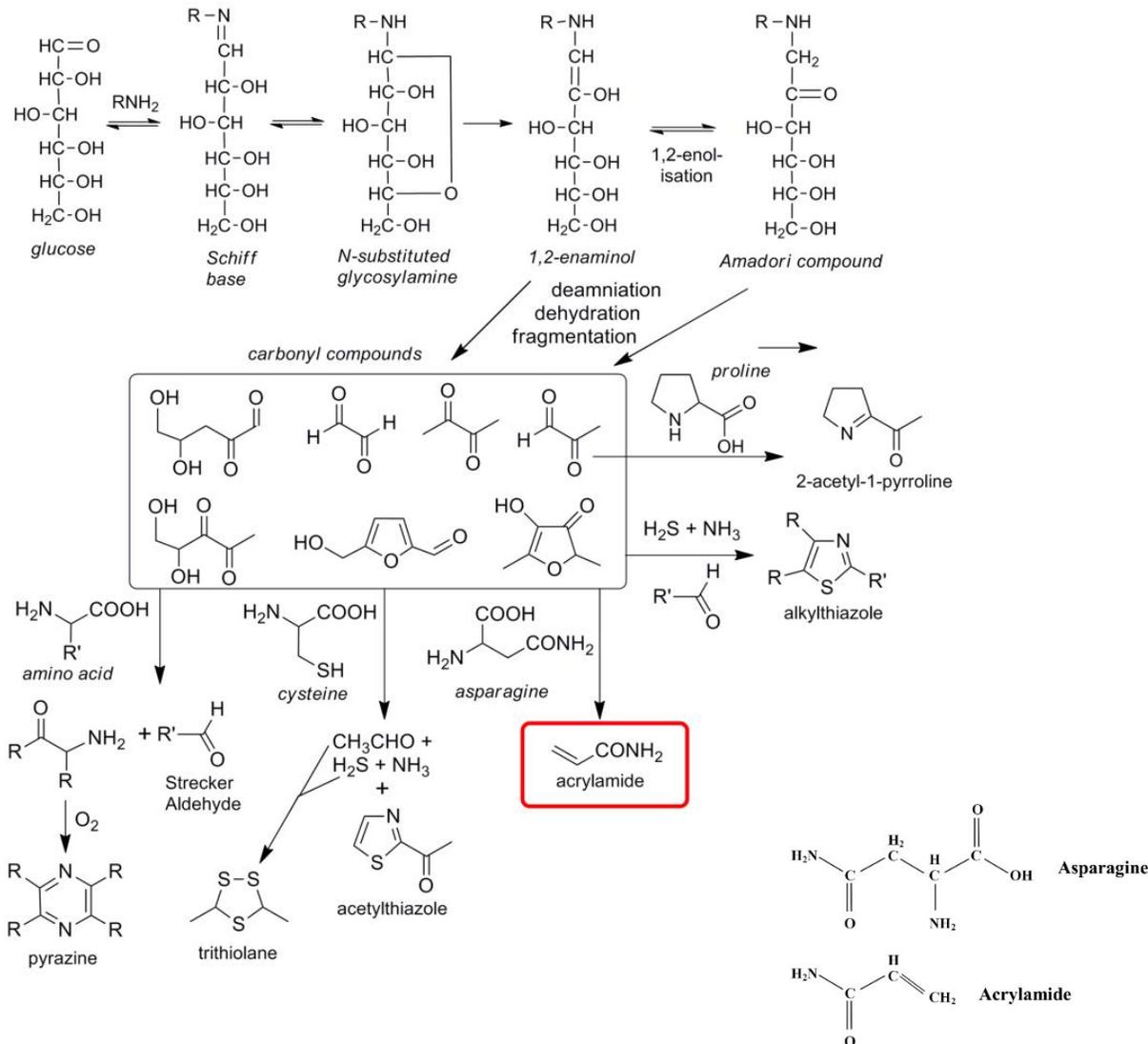


- All food businesses to monitor the levels of acrylamide in their products and to keep a record of the mitigation measures they apply
- Compulsory mitigation measures (Codes of Practice), many of which reflect the FoodDrinkEurope Acrylamide Toolbox
- Benchmark Levels set for different food products, replacing Indicative Values.
- Mitigation measures to be reviewed if the levels of acrylamide in a product are not below the Benchmark Level.
- Threat of Maximum Levels with regulatory enforcement for sectors not showing sufficient progress: **the European Commission has already started this process.**

Acrylamide forms in the Maillard reaction



ROTHAMSTED
RESEARCH



Free amino acids react with reducing sugars (glucose, fructose and maltose) at high temperature in the Maillard reaction to produce a plethora of flavour, aroma and colour compounds.

Acrylamide is formed when free asparagine participates in the final stages of the reaction.

This is the predominant route for the generation of acrylamide during cooking.

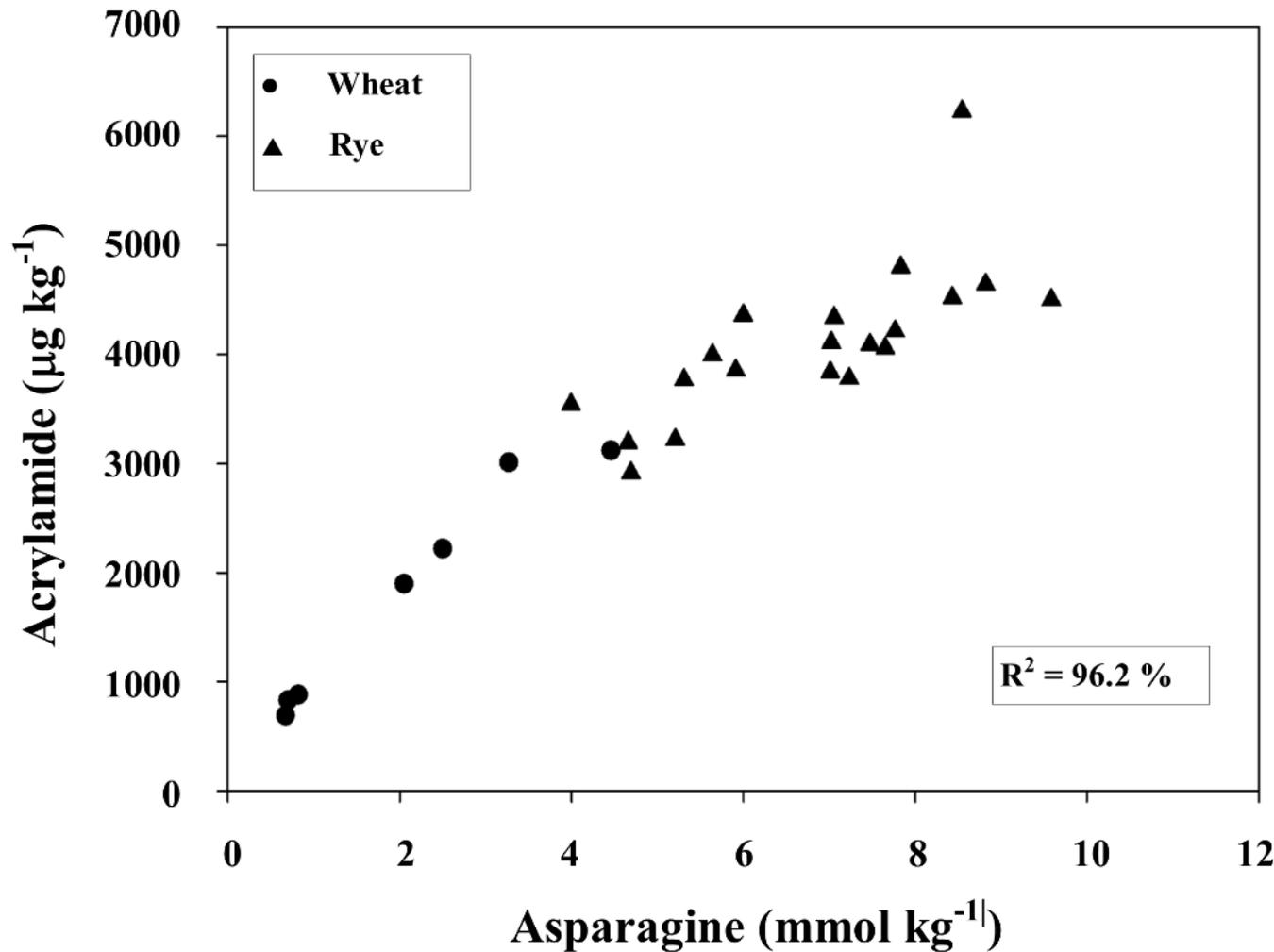
Free (soluble, non-protein) asparagine and reducing sugars can therefore be regarded as precursors of acrylamide, but other free amino acids can play a part, positively and negatively.



Free (non-protein) asparagine concentration determines acrylamide formation in heated flour produced from wheat and rye grain



ROTHAMSTED
RESEARCH



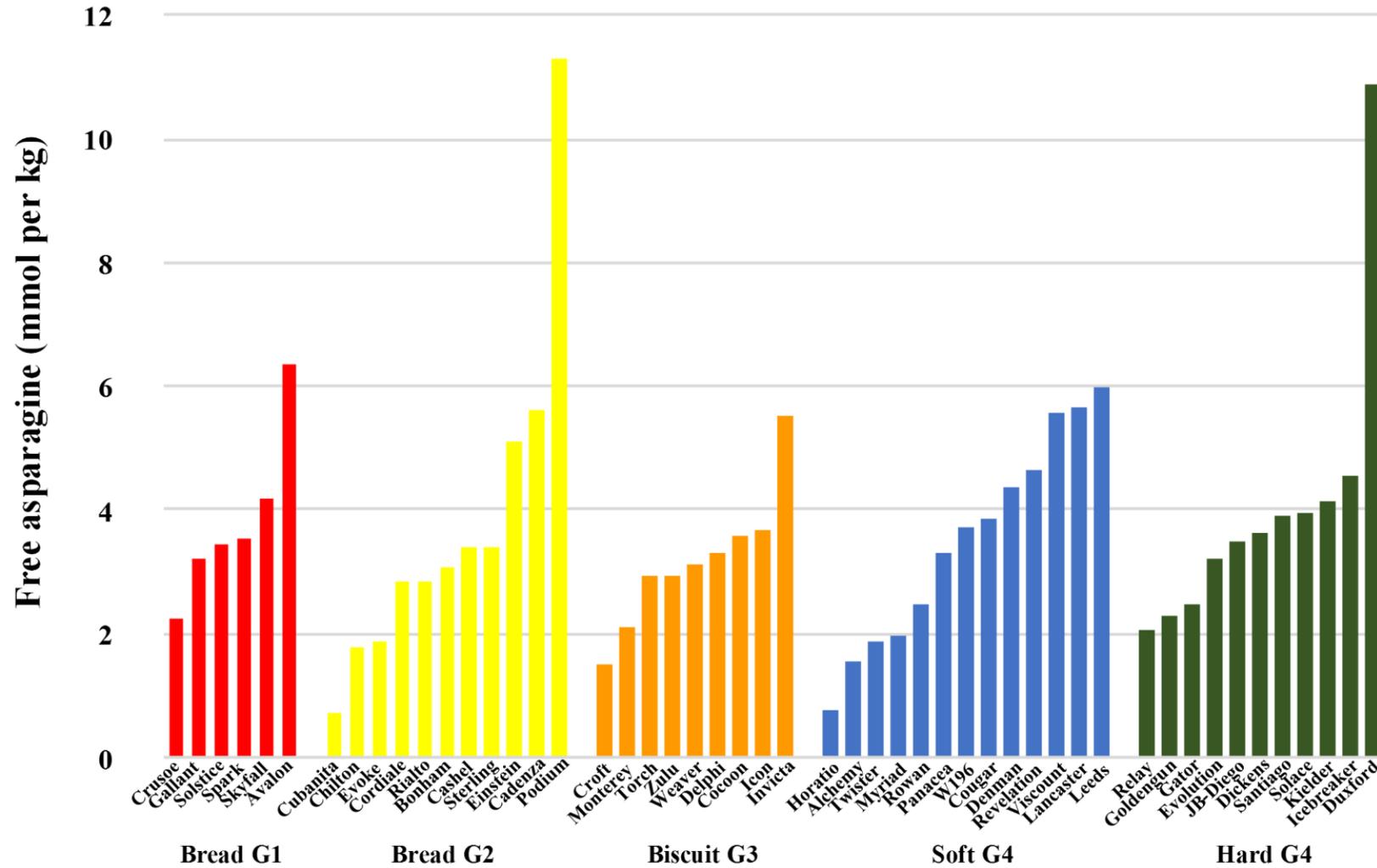
Acrylamide formation ($\mu\text{g}/\text{kg}$) plotted against free asparagine concentration (mmol/kg) in wheat and rye flour heated at $180\text{ }^{\circ}\text{C}$.

A consideration of free asparagine concentration in the raw material forms part of the cereal products Code of Practice.

Varietal differences in wheat



ROTHAMSTED
RESEARCH

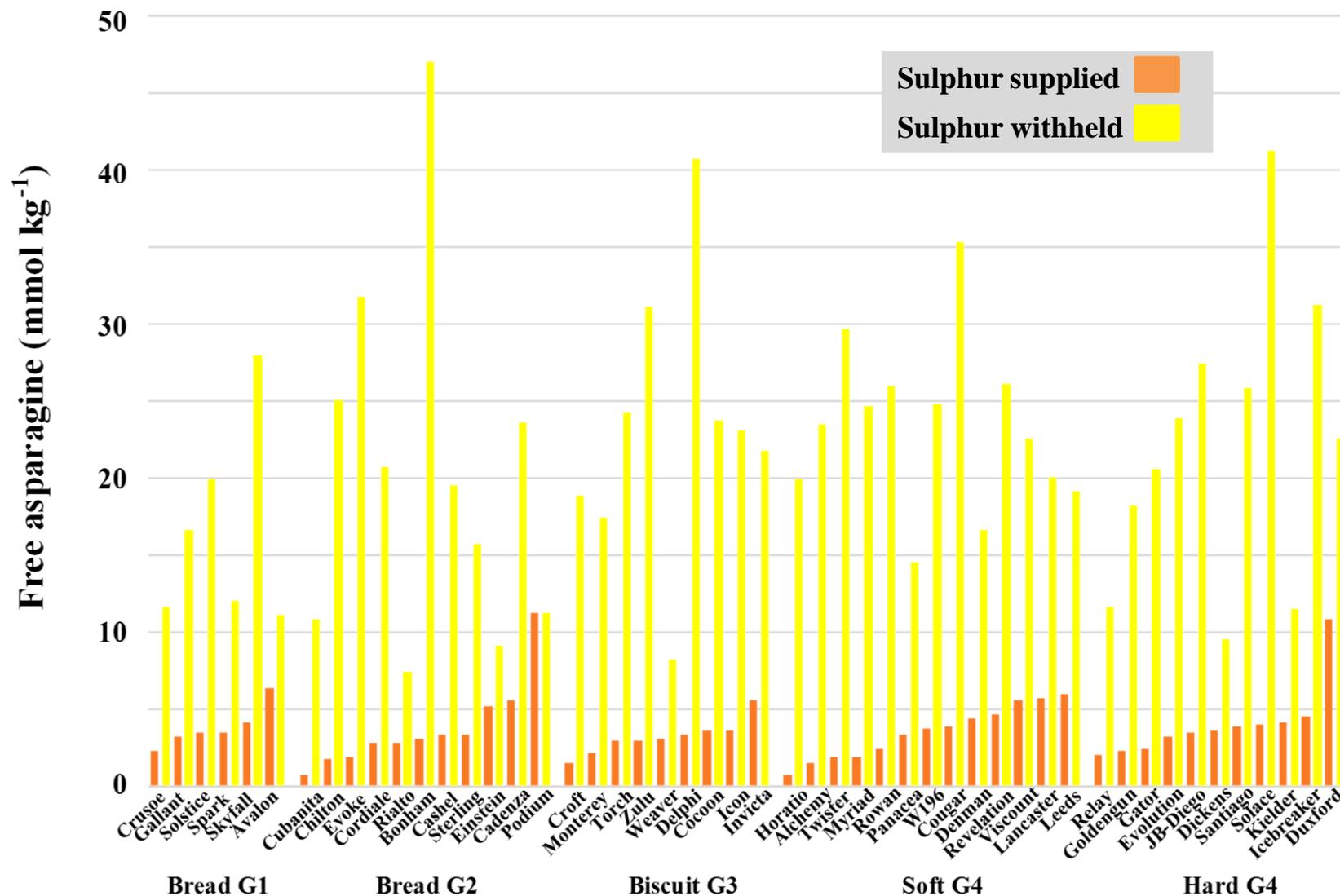


Free asparagine concentration in grain of winter wheat varieties from the UK's AHDB Recommended List grown in a field trial in the UK

Crop management: the importance of supplying wheat with sufficient sulphur



ROTHAMSTED
RESEARCH



Mean free asparagine concentration in the grain of 50 varieties of winter wheat grown in a field trial in 2012-2013. Data are shown from split-plots in which wheat in half the plot was supplied with nitrogen and sulphur (brown bars), while the wheat in the other half was supplied with nitrogen but not sulphur (yellow bars).

Ensuring sulphur sufficiency is also included in the European Commission's compulsory Code of Practice for cereals.

Field trials aimed at refining our advice on N and S application and other minerals



ROTHAMSTED
RESEARCH



- Woburn UK, 2019-2022
- Different ratios of N and S
- Also looking at phosphate and potassium

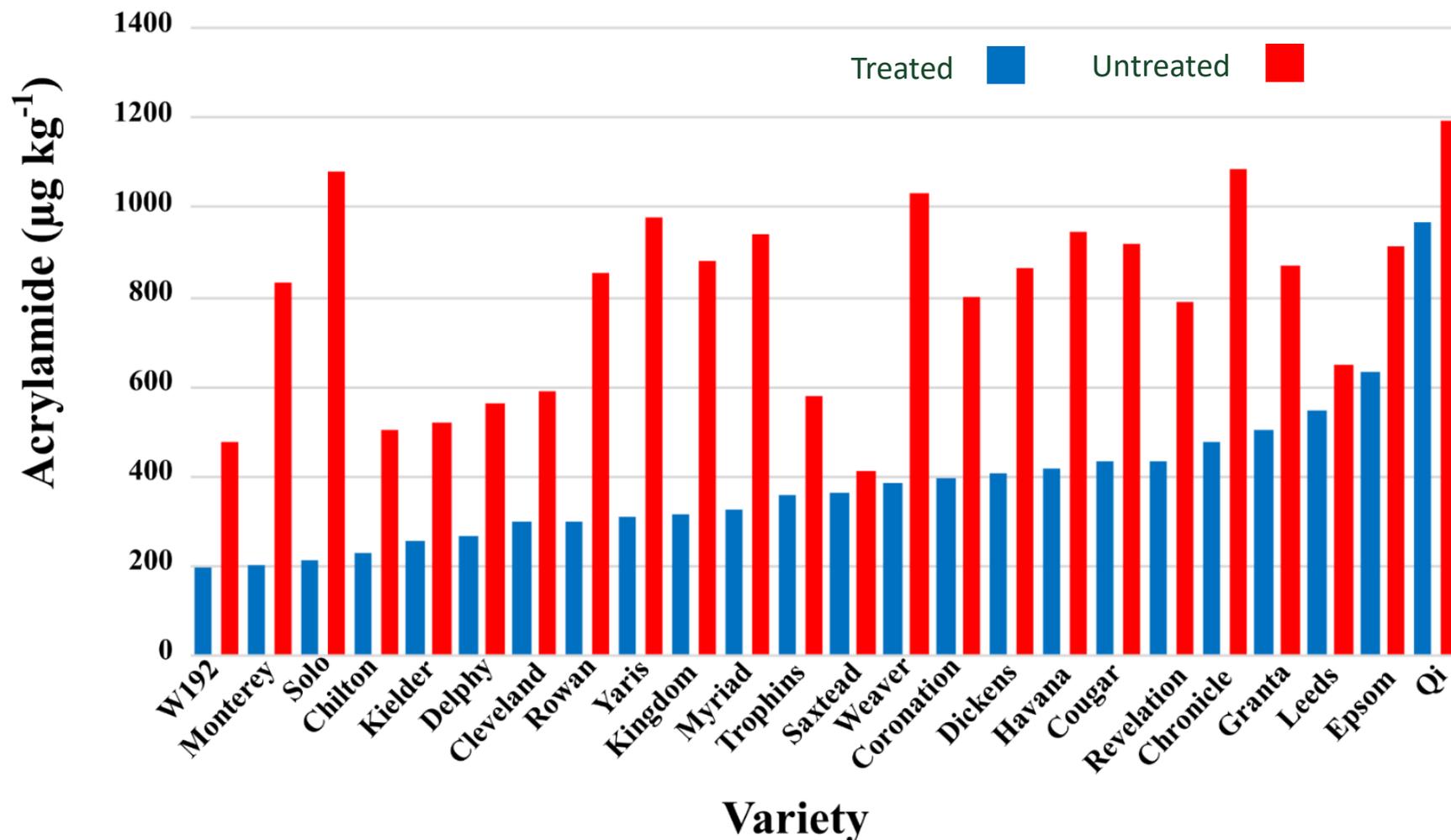
See Joe Oddy's poster.



Crop management: Acrylamide formation in flour from wheat grown with or without fungicide treatment



ROTHAMSTED
RESEARCH



Untreated plots showed visible infection by *Septoria tritici*, Yellow rust and Brown rust.

To follow 'good phytosanitary practices to prevent fungal infection' has been included in the European Commission's compulsory Code of Practice for wheat products

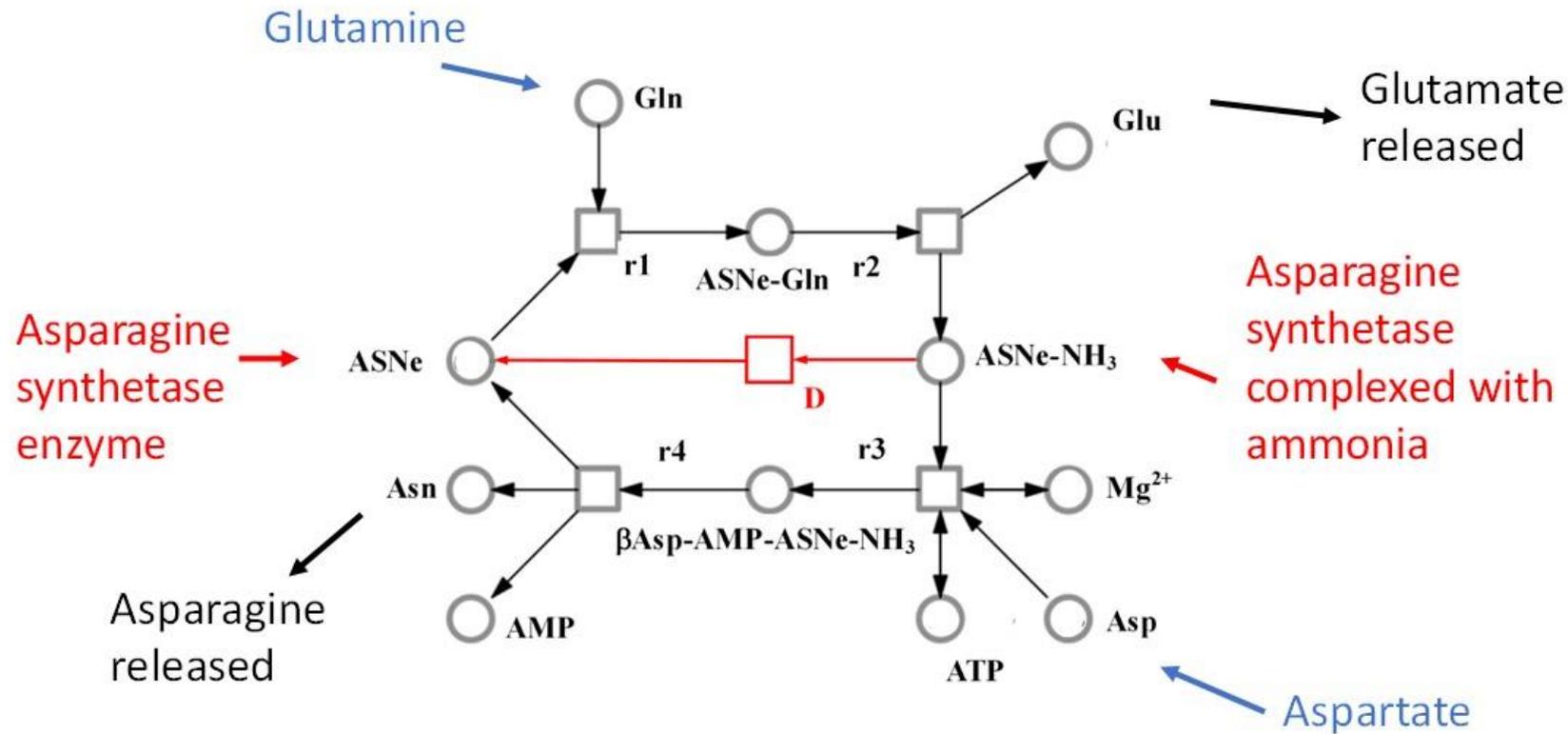


- To achieve a step reduction in free asparagine concentration in wheat grain without affecting other quality traits, yield, disease resistance etc.
- Longer term:
To uncouple asparagine concentration from the effects of nutrition (N and S) and pathogens.
To re-engineer other aspects of amino acid metabolism in wheat grain for biofortification, using low asparagine wheat as a starting point.

Genetic targets: asparagine synthetase



ROTHAMSTED
RESEARCH



Asparagine synthetase catalyses the ATP-dependent transfer of an amino group from glutamine to aspartate to produce glutamate and asparagine.

Genetic targets: Wheat asparagine synthetase genes



ROTHAMSTED
RESEARCH

TaASN1 on Chromosome 5



TaASN2 on Chromosome 3



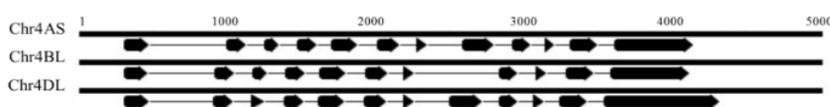
TaASN3.1 on Chromosome 1



TaASN3.2 on Chromosome 1



TaASN4 on Chromosome 4



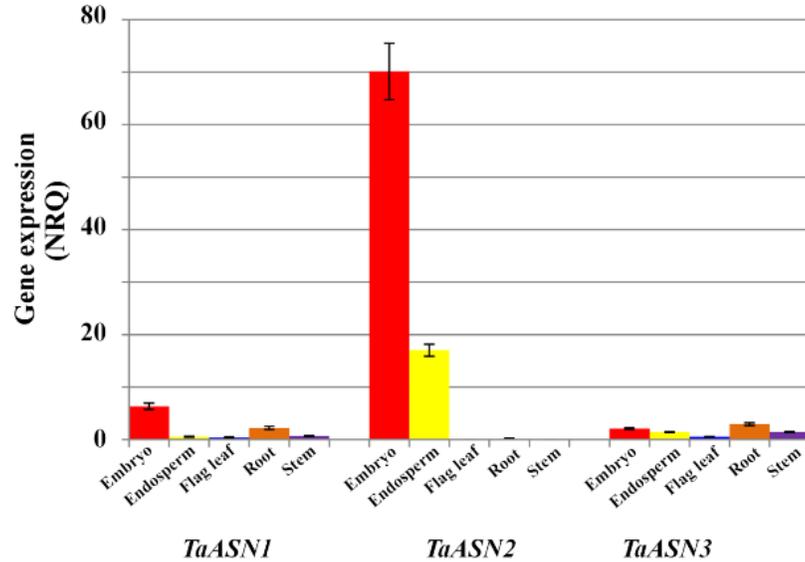
Key: Exon Intron

- Five genes per genome: *TaASN1*, *TaASN2*, *TaASN3.1*, *TaASN3.2* and *TaASN4*.
- Each gene is present as a single copy.
- Some varieties lack a *TaASN2* gene on Chromosome 3B.
- This gene family structure is unique to the Triticeae.

Differential expression of asparagine synthetase genes in different tissues of wheat

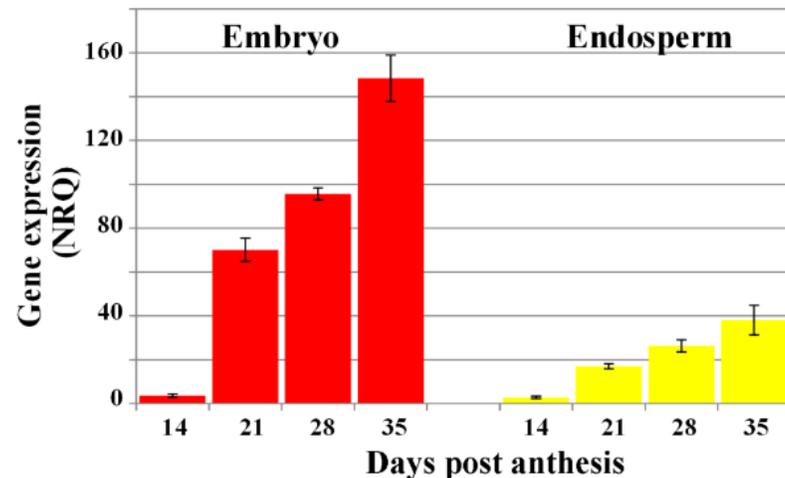


ROTHAMSTED
RESEARCH



Expression (NRQ means and standard errors) of *TaASN1*, *TaASN2* and *TaASN3* in different tissues of wheat.

TaASN2 is an obvious target for genetic intervention

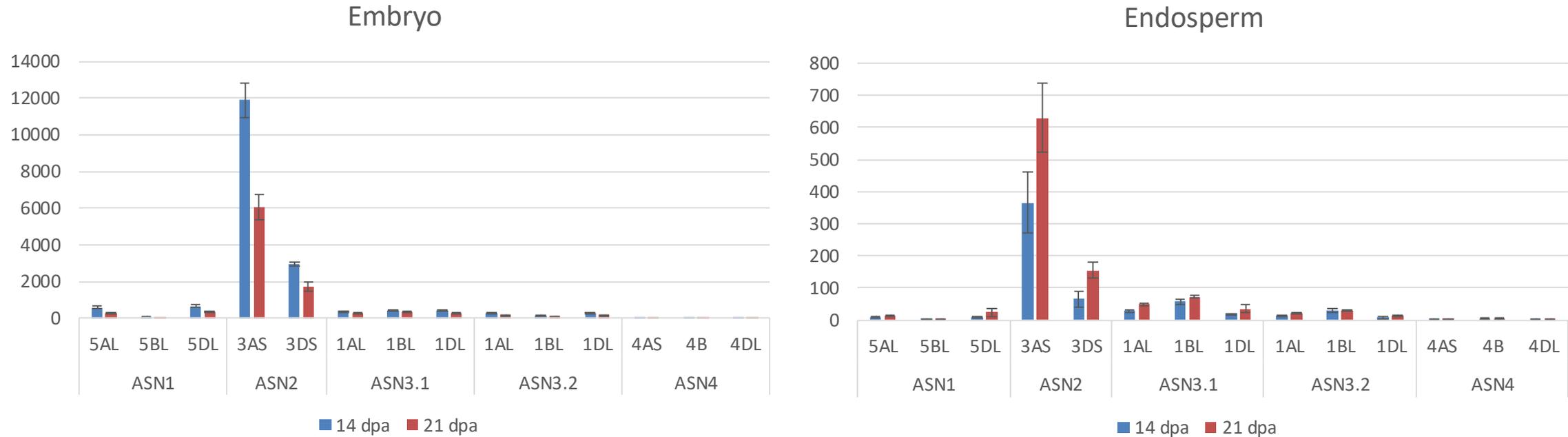


Expression of *TaASN2* in the embryo (bran fraction) and endosperm (white flour fraction) through grain development.

Asparagine synthetase gene expression: RNA-seq data



ROTHAMSTED
RESEARCH

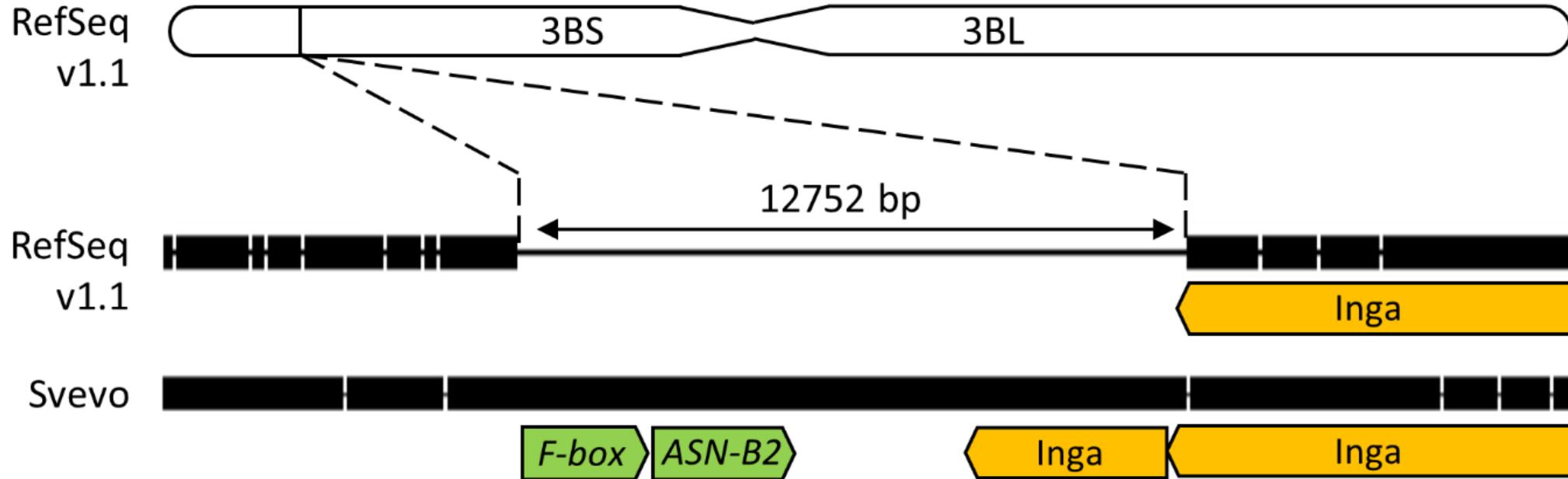


- Higher levels of expression (> 10-fold difference) in total asparagine synthetase gene expression in the embryo than in the endosperm.
- *TaASN2* was the most highly expressed in both tissues.
- The A genome *TaASN2* gene was much more highly expressed (> 3-fold difference) than the D genome copy. The *TaASN-B2* gene is absent in these genotypes.
- Reaffirmed *TaASN2* as the best target! Knocking out the *TaASN-A2* gene on its own may be effective.

A natural deletion in Chromosome 3B of variety Chinese Spring includes the *TaASN-B2* gene



ROTHAMSTED
RESEARCH



Alignment of the region of Chromosome 3B of variety Chinese Spring where the *TaASN-B2* gene has been deleted, and alignment with the corresponding region in variety Svevo, in which the *TaASN-B2* gene is present.

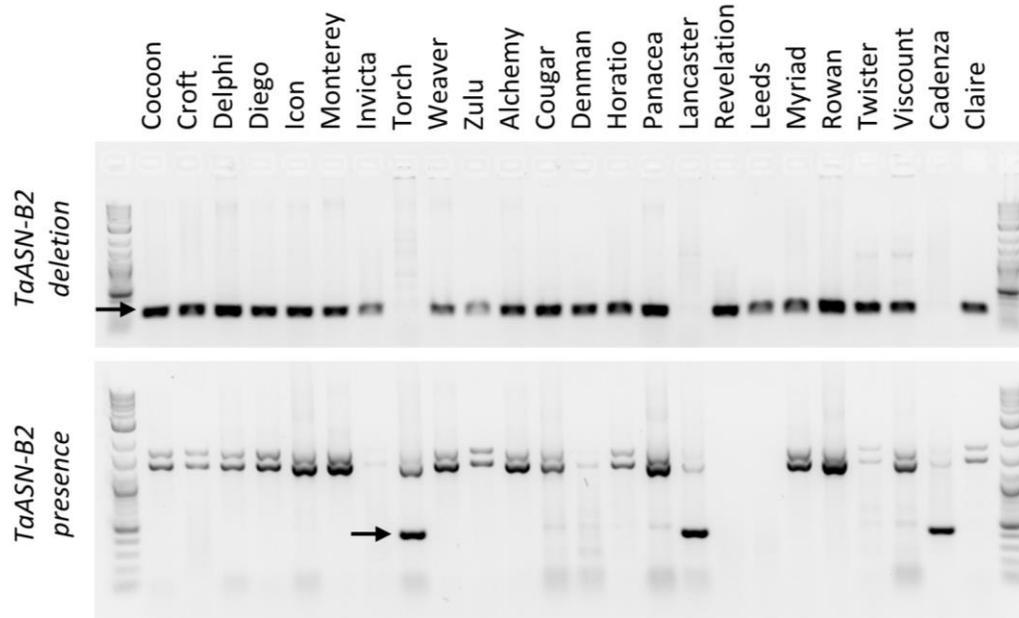


Joe Oddy

Screening for the presence/absence of the *TaASN-B2* gene in UK wheat varieties

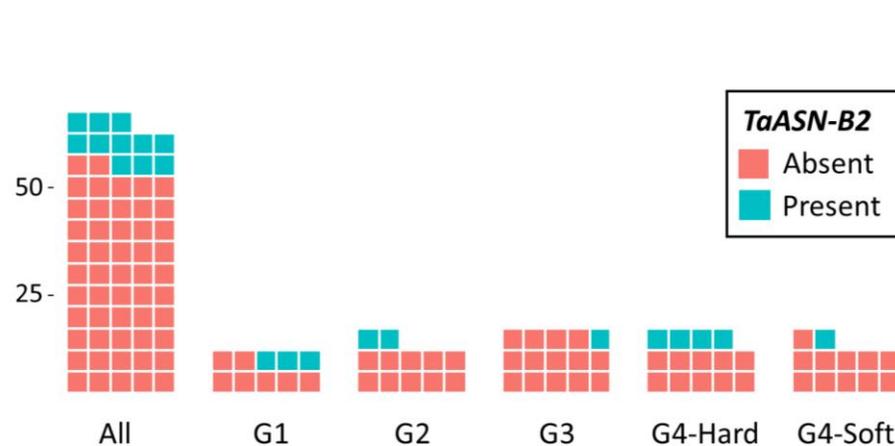


ROTHAMSTED
RESEARCH



Top. Example of an electrophoresis gel of PCR products from assays to distinguish the presence and absence of *TaASN-B2* in a collection of UK wheat varieties. Varieties Cadanza and Claire were used as controls for *TaASN-B2* presence and absence, respectively.

- Most but not all UK wheat varieties lack a *TaASN-B2* gene.
- The deletion is present in some but not all wild wheat relatives.

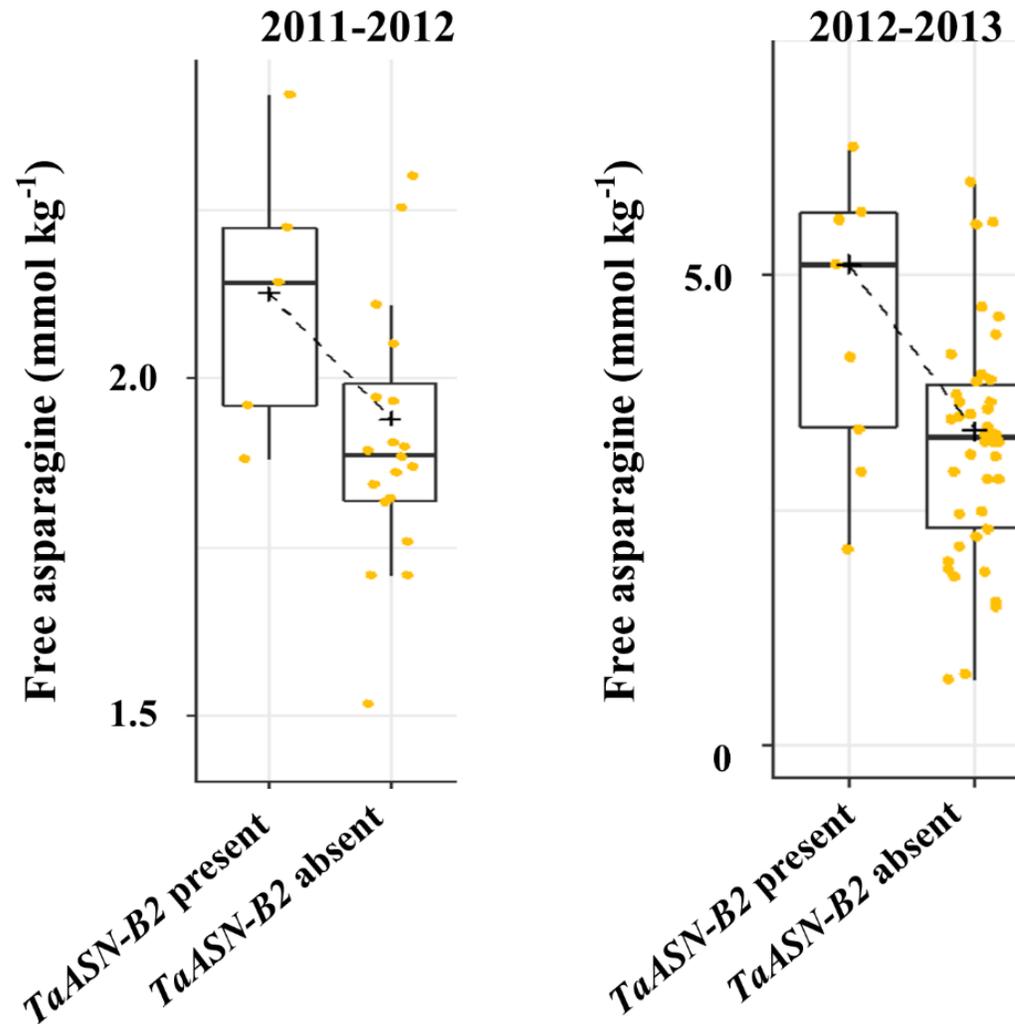


Bottom. Diagram showing the frequency of the *TaASN-B2* deletion in 63 UK wheat varieties, separated into UK Flour milling groups: G1 (breadmaking), G2 (breadmaking potential), G3 (soft/biscuit), G4 (feed/other).

Effect of the *TaASN-B2* deletion on grain free asparagine levels in two year's of field trials



ROTHAMSTED
RESEARCH



- The presence of the *TaASN-B2* gene was associated with higher free asparagine concentrations.
- This breaks down under sulphur deficiency (not shown).
- See Joe Oddy poster for more on this and work to identify QTL for asparagine concentration

Knocking out *TaASN2* to produce low asparagine wheat: Genome editing with CRISPR-Cas9



ROTHAMSTED
RESEARCH

Four gRNAs were designed to target the first exon of *TaASN2*

1F -> GGGGTGCGGCGACGAGTCGCAGG

1 ATGTGCGGCATACTAGCGGTGCTGGGGTTCGCGGCGACGAGTCGCAGGGAAGAGGGTCCACGTGCTAGAGCTCTCGCGCAG
1 M **C** G I L A V L G C G D E S Q G K R V H V L E L S R R

2F -> GGACTGGAGCGGCCTGCACCAGG

81 GCTCAAGCACCGGGCCCGGACTGGAGCGGCCTGCACCAGGTCGCCGACAACACTACCTCTGCCACCAGCGCCTCGCCATCA
28 L K H R G P D W S G L H Q V A **D** N Y L C H Q **R L A I** I

GGAGCCGCTGGTCGGCGAGATG <- 3R

161 TCGACCCGGCCTCCGGCGACCAGCCGCTCTACAACGAGGACAAGTCCATCGCCGTTGCCGTC AACGGGGAGGTCTACAAC
55 D P A S G D Q P L Y N E D K S I A V A V **N G E** V Y N

GGCCTGGCCGTCACTGACGCTCC <- 4R

241 CATGAGGAGCTTCGGGCACGGCTCTCCGGACACAGGTTCCGGACCGGCAGTGACTGCGAGGTCATCGCCCATCTG
81 H E E L R A R L S G H R F R T G S **D** C E V I A **H** L



Sarah Raffan

A gene encoding the 4 gRNAs was introduced into wheat by particle bombardment, along with a gene encoding the Cas9 nuclease and a selective marker gene (*Bar*).

Types of editing induced in our wheat plants by CRISPR



ROTHAMSTED
RESEARCH

```

20 1F                               95 2F
WT  TGCTGGGGTGCGGCGACGAG-TCGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
L23 A1 TGCTGGGGTGCGGCGACGAG-TCGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
L23 A2 TGCTGGGGTGCGGCGACGAG-TCGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
L23 B1 TGCTGGGGTGCGGCGACGAG-TCGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
L23 B2 TGCTGGGGTGCGGCGACGAG-TCGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
L23 D1 TGCTGGGGTGCGGCGACGAG-TCGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
    
```

Line 23: Total null

Most of the edits are deletions, but there are substitutions and single base insertions as well.

```

L59 A7 TGCTGGGGTGC---CAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
L59 A8 TGCTGGGGTGCGGCGACGAG-TCGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
L59 B5 TGCTGGGGTGCGGCGACGAG-TCGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
L59 D4 TGCTGGGGTGCGGCGAC---CGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
    
```

Line 59: Total null

```

L178 A11 TGCTGGGGTGCGGCGACGAG-TCGCAGGGGAAGAGGGTCCA...GCCC GGACTGGAGCGGCTGCA-CCAGGTCGCCGACAACCTCTGCCACCAGCGC...
    
```

Line 178: A genome null

Most of these edited genes encode highly truncated, dysfunctional proteins.

```

157 3R                               266 4R
WT  ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
L23 A1 ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
L23 A2 ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
L23 B1 -----CGGCAGTGACTGCGAGGTCAT...
L23 B2 ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
L23 D1 ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
    
```

Line 23: Total null

Line 59 also has an edit in the *TaASN-B1* gene; i.e. it is a partial knockout for *TaASN1*: see Sarah Raffan's poster

```

L59 A7 ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
L59 A8 ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
L59 B5 ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
L59 D4 ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
    
```

Line 59: Total null

```

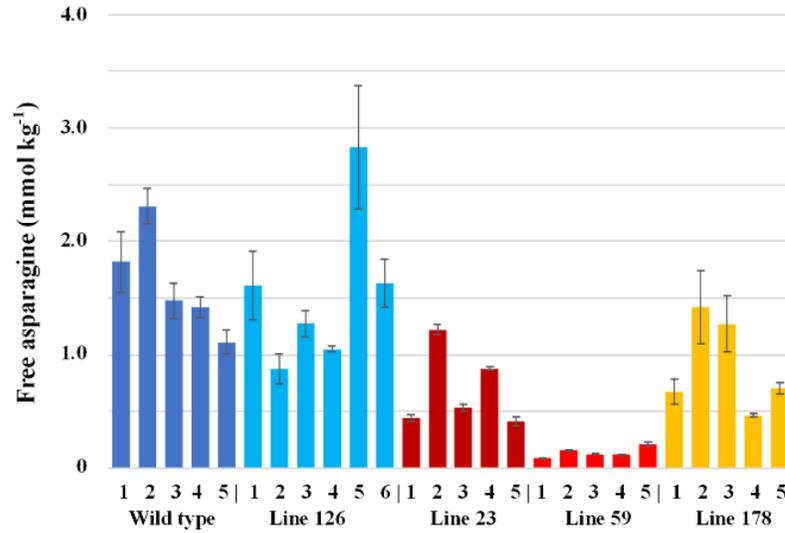
L178 A11 ATCATCGACCCGGCCTCCG-GCGACCAGCCGCTCTACAACG...CCGGACACAGGTTCCGGAC-CGGCAGTGACTGCGAGGTCAT...
    
```

Line 178: A genome null

Free asparagine levels in grain of control and edited *TaASN2* knockout lines



ROTHAMSTED
RESEARCH



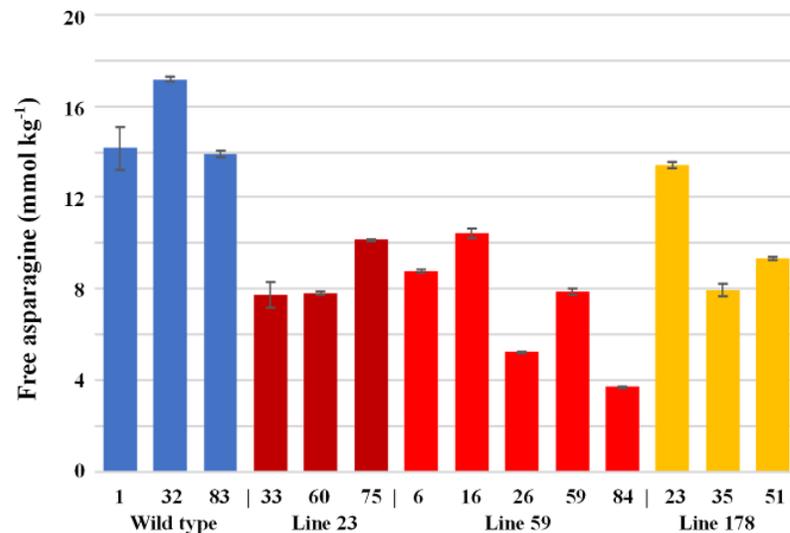
T2 top and T3 (bottom) seed. Free asparagine and total free amino acid levels were higher in the T3 generation, possibly due to stress.

Lines 23 and 59 are total knockouts

Line 178 is a partial knockout

Line 126 is not edited.

Line 59 is also a partial *TaASN1* knockout.

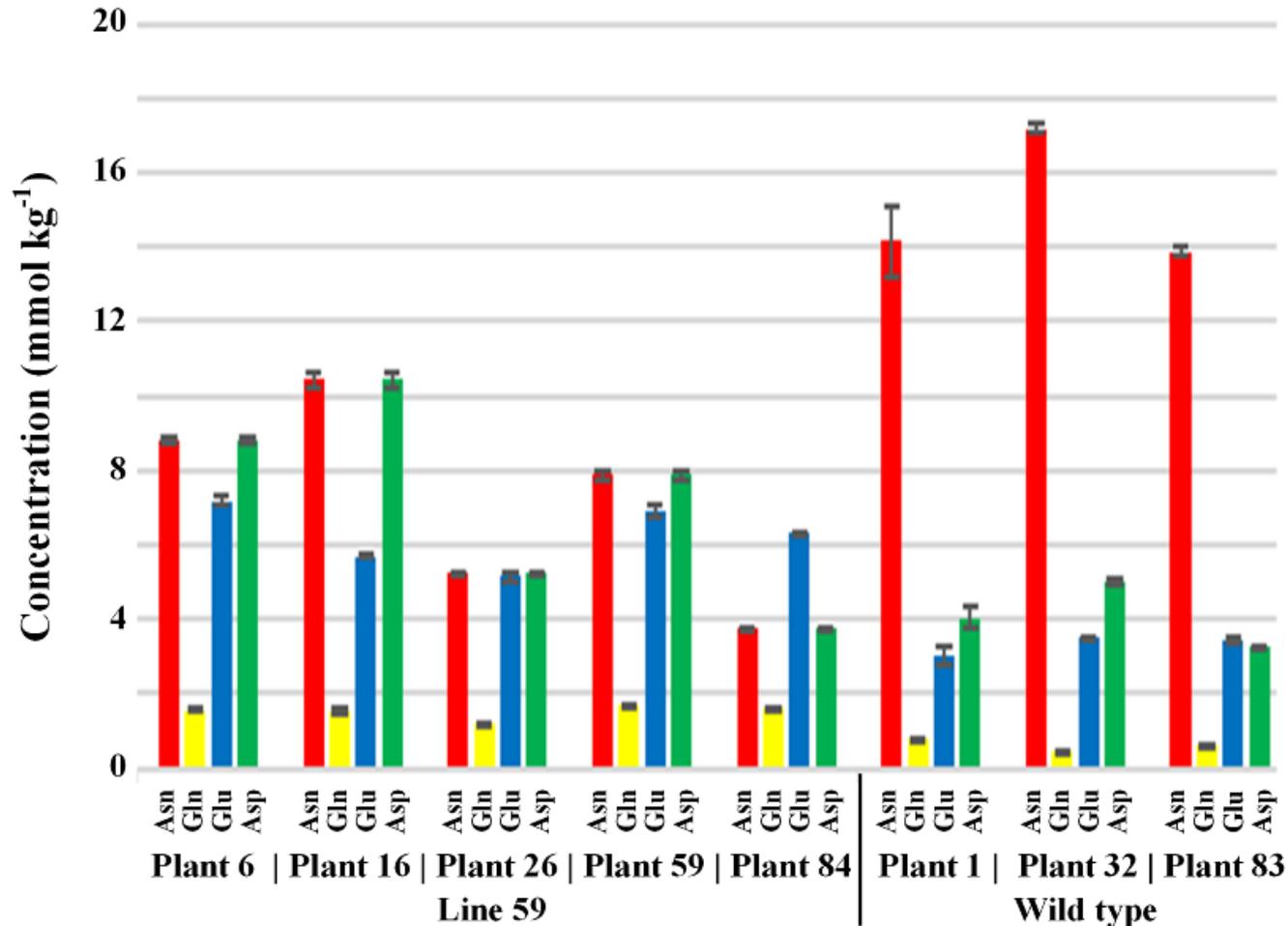


Currently they still contain some transgenes but these are being removed by selfing and crossing and we now have some plants from line 59 that are transgene-free.

The ratios of free asparagine to free glutamine, glutamate and aspartate are altered in the CRISPR lines



ROTHAMSTED
RESEARCH



The graph shows the means and standard errors of free asparagine, glutamine, glutamate and aspartate concentrations in wholemeal flour prepared from the grain of edited wheat plants of line 59 and wildtype controls.

CRISPR editing of *TaASN2*



ROTHAMSTED
RESEARCH

- Our study achieved a high mutation rate, with 11 lines derived from 14 selected T0 plants showing editing. We put this down to the simultaneous introduction of four gRNAs along with a wheat-optimised *Cas9* gene.
- It required NGS analysis to characterise the edits in the plants.
- Editing continued beyond the T0 generation, but only in some of the lines, and we also saw evidence of somatic editing. We conclude that several generations and segregating away of the *Cas9* gene may be required to achieve stability.
- Most of the edits were deletions, the longest of which was of 173 base pairs. However, there were some insertions as well, all of a single base pair, and some substitutions.
- The wheat asparagine synthetase gene family lent itself to the aims of the study. Non-Triticeae cereals, such as maize and rice, do not have an equivalent of the *TaASN2* gene.
- Poor germination was seen in some of the low asparagine lines but this could be overcome by application of asparagine and was not seen in every generation. This requires further investigation.

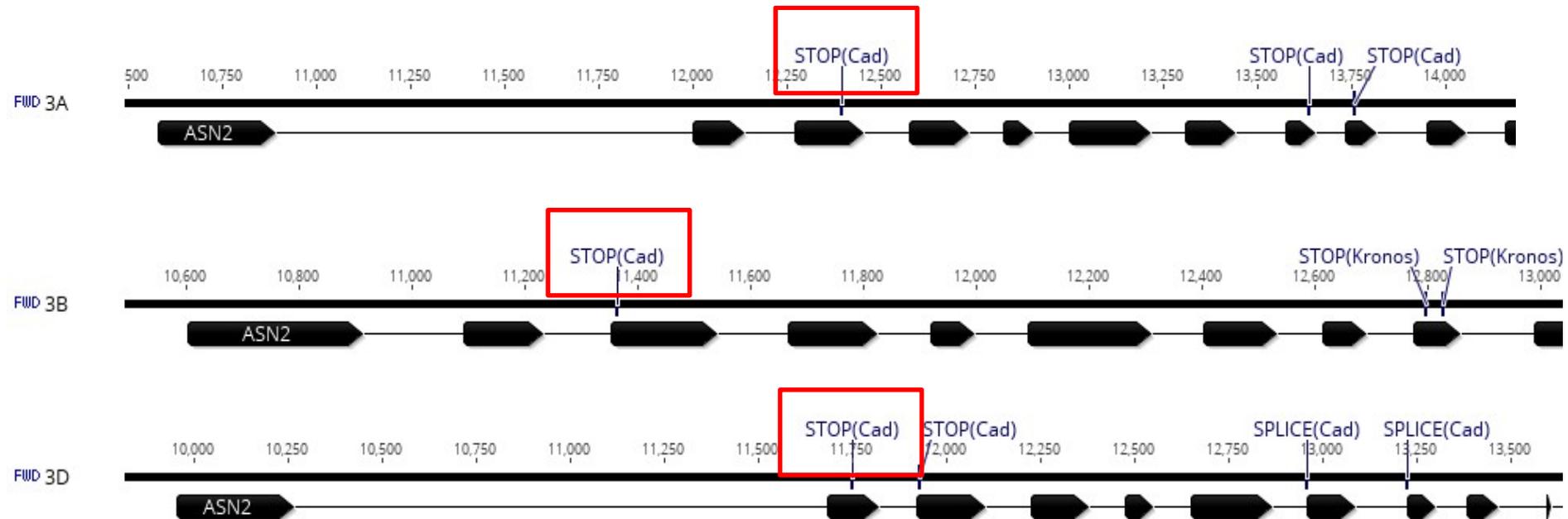


Knocking out *TaASN2*: stacking mutations from TILLING lines



ROTHAMSTED
RESEARCH

- Target Induced Local Lesions In Genome (TILLING)
- Two EMS-mutagenised populations, in tetraploid durum wheat cv 'Kronos' and hexaploid bread wheat cv 'Cadenza', which have been exome-sequenced and characterised
- Developed as part of a joint project between the University of California Davis, Rothamsted Research, The Earlham Institute, and John Innes Centre
- Lines have been identified carrying mutations in the *TaASN2* genes in each of the A, B and D genomes
- The mutations are being stacked in variety Claire by our partners at RAGT to generate a null *TaASN2* line (Claire already lacks *TaASN-B2*).



Europe's first CRISPR wheat field trial



**ROTHAMSTED
RESEARCH**



Department
for Environment
Food & Rural Affairs

Seacole Building
2 Marsham Street
London SW1P 4DF

Victoria Prentis MP
Parliamentary Under Secretary of State

T 03459 335577
deira.hejlskov@defra.gov.uk
www.gov.uk/defra

Professor Nigel Halford
Rothamsted Research
West Common
Harpenden
Hertfordshire
AL5 2JQ

Our ref: 21/R08/01

3rd August 2021

ENVIRONMENTAL PROTECTION ACT 1990, SECTIONS 111 AND 112:

CONSENT TO RELEASE GENETICALLY MODIFIED ORGANISMS REFERENCE 21/R08/01

1. In accordance with section 111 of the Environmental Protection Act 1990, the Secretary of State for Environment, Food and Rural Affairs hereby grants consent to Rothamsted Research to perform the release of the genetically modified organisms described in paragraph 2, in accordance with the particulars set out in paragraph 3, and subject to the limitations and conditions set out in the Schedule attached.

2. Genetically Modified Organism to be released:

The genetically modified organisms (GMOs) are wheat *Triticum aestivum* plants that have been gene edited using CRISPR-Cas9 to contain point mutations in the TaASN2 asparagine synthetase gene.

3. Particulars of the consent to release:

Maximum size of the release: The area sown with the GM wheat must not exceed 1500 square metres and may take the form of one or more plots within the trial site.

(a) Purpose of the release:

To investigate the effect of knocking out the TaASN2 gene on free asparagine accumulation in wheat grain in the field.

(b) Location of the release ("trial site"):

Britain to grow cancer-cutting wheat for making healthier bread

British scientists will grow a new strain of wheat using a revolutionary form of genetic editing as part of an effort to make healthier bread.

The trials will be the first in Europe for a wheat that has been genetically edited using a tool known as Crispr. It comes as the government consults on whether to use post-Brexit freedom to break with European Union rules that have limited the use of the technology in agriculture.

The trial, which had to be approved by the government, will be run by Rothamsted Research in Hertfordshire, which will grow the crop outside. The grain has been gene-edited to reduce levels of asparagine, a naturally occurring amino acid. When wheat is used to make bread and food, asparagine is converted into acrylamide, which is thought to be carcinogenic.

Professor Nigel Halford, leading the project, said the aim was to produce healthier wheat without changing the taste, that would not be considered to be genetically modified (GM). GM was originally used as a label for crops where genes had been transferred from one organism into another, sometimes across non-related species, in a way that does not happen naturally.

The Crispr tool enables the genetic material of a plant to be precisely edited without new material being added.

Scientists have argued that Crispr is fundamentally different from conventional GM, in part because the changes that result could occur through natural mutation.

In 2018 the EU Court of Justice ruled that Crispr-edited crops should be subject to the same stringent regulations as conventional GM organisms, which some scientists said would slow research.

For the wheat in Hertfordshire, GM is used as a label for the wheat.

Continued on page 2, col 3

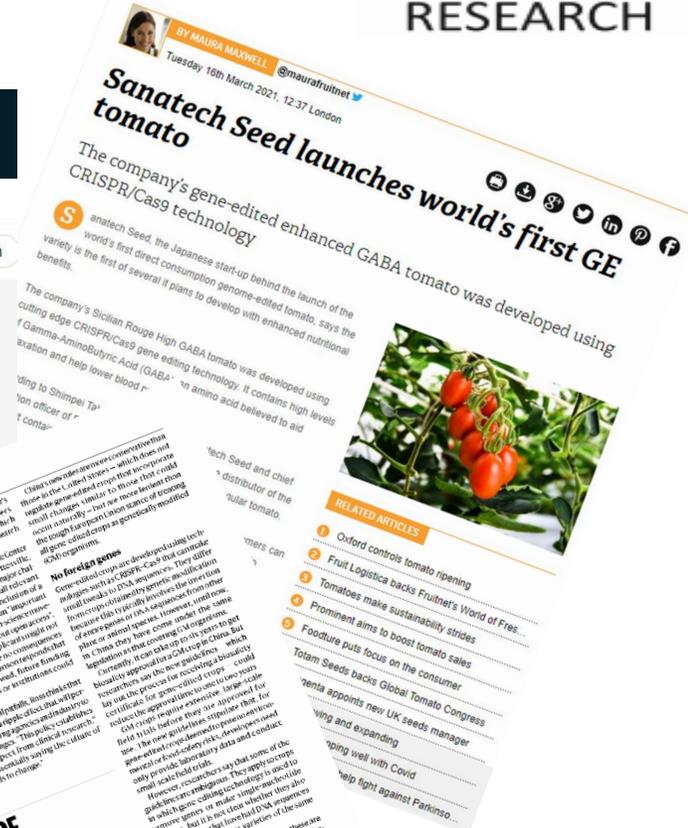


Regulation of genome edited crops



ROTHAMSTED RESEARCH

- The EU currently has no formal regulations on the genome editing of crops, but as a result of European Court of Justice ruling on case C-528/16 (25th July 2018), gene edited plants have to be treated like transgenic plants, and edited genes as if they were transgenes.
- Many other regulatory authorities around the world have taken a very different view (USA, Brazil, Argentina, Japan, Canada, Chile, Colombia, Israel, China, India for example).

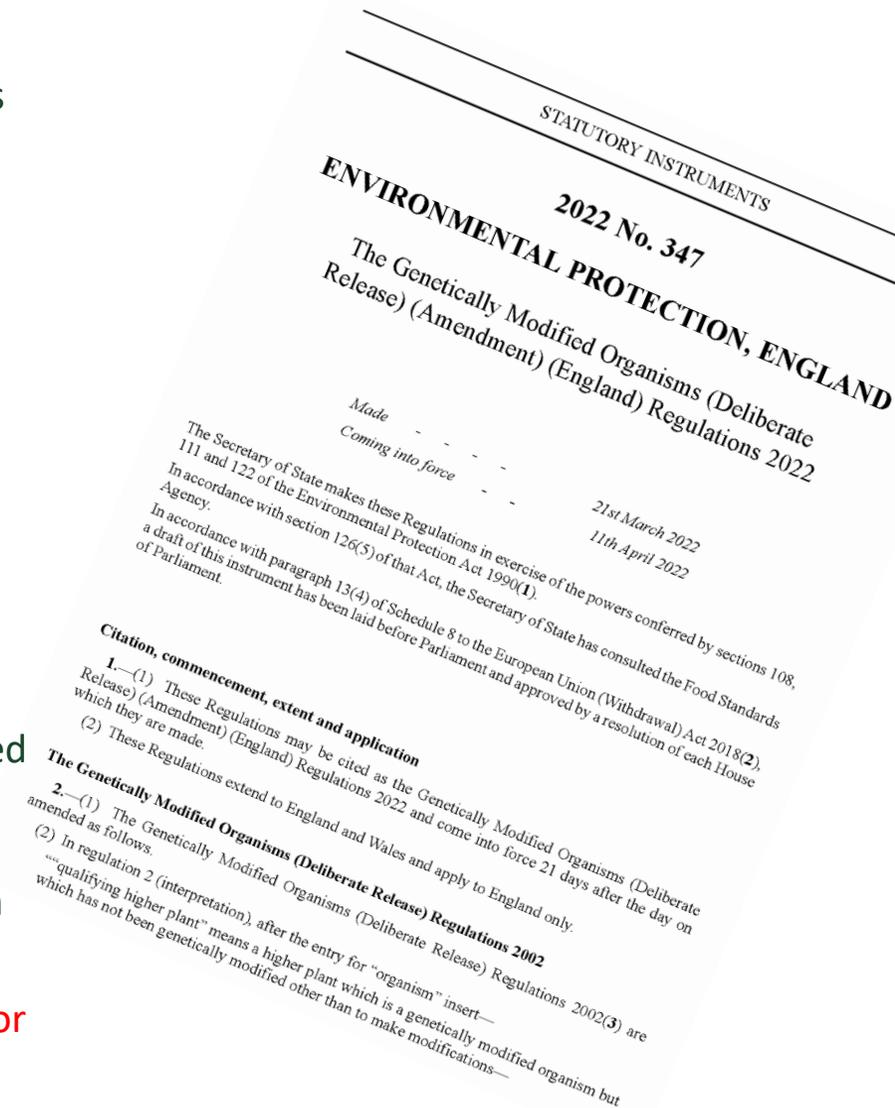


UK: Changes to regulations on genome editing



ROTHAMSTED
RESEARCH

- The UK currently operates under legacy regulations from the EU.
- UK Government announcement, 29th September 2021: Plans to unlock potential benefits of gene editing set out.
- As a first step the government will change the rules relating to gene editing to cut red tape and make research and development easier.
- Research scientists will still need to notify Defra of any trials involving GE plants.
- This first step applies to research and development only; for any products that are authorised for market, existing GMO rules would apply.
- The government will also consider the appropriate measures needed to enable gene edited products to be brought to market safely and responsibly.
- In the longer term, this will be followed by a review of its approach to GMO regulation more broadly.
- The Statutory Instrument (SI) on removing GE field trials from GMO regulation was passed last month and comes into effect today: The Genetically Modified Organisms (Deliberate Release) (Amendment) (England) Regulations 2022 (legislation.gov.uk/ukxi/2022/347/regulation/2/made), so things are already in motion in England.
- **Probably the first positive step in the regulation of agricultural biotechnology in the UK for 25 years.**



UK: Changes to regulations on genome editing



ROTHAMSTED
RESEARCH

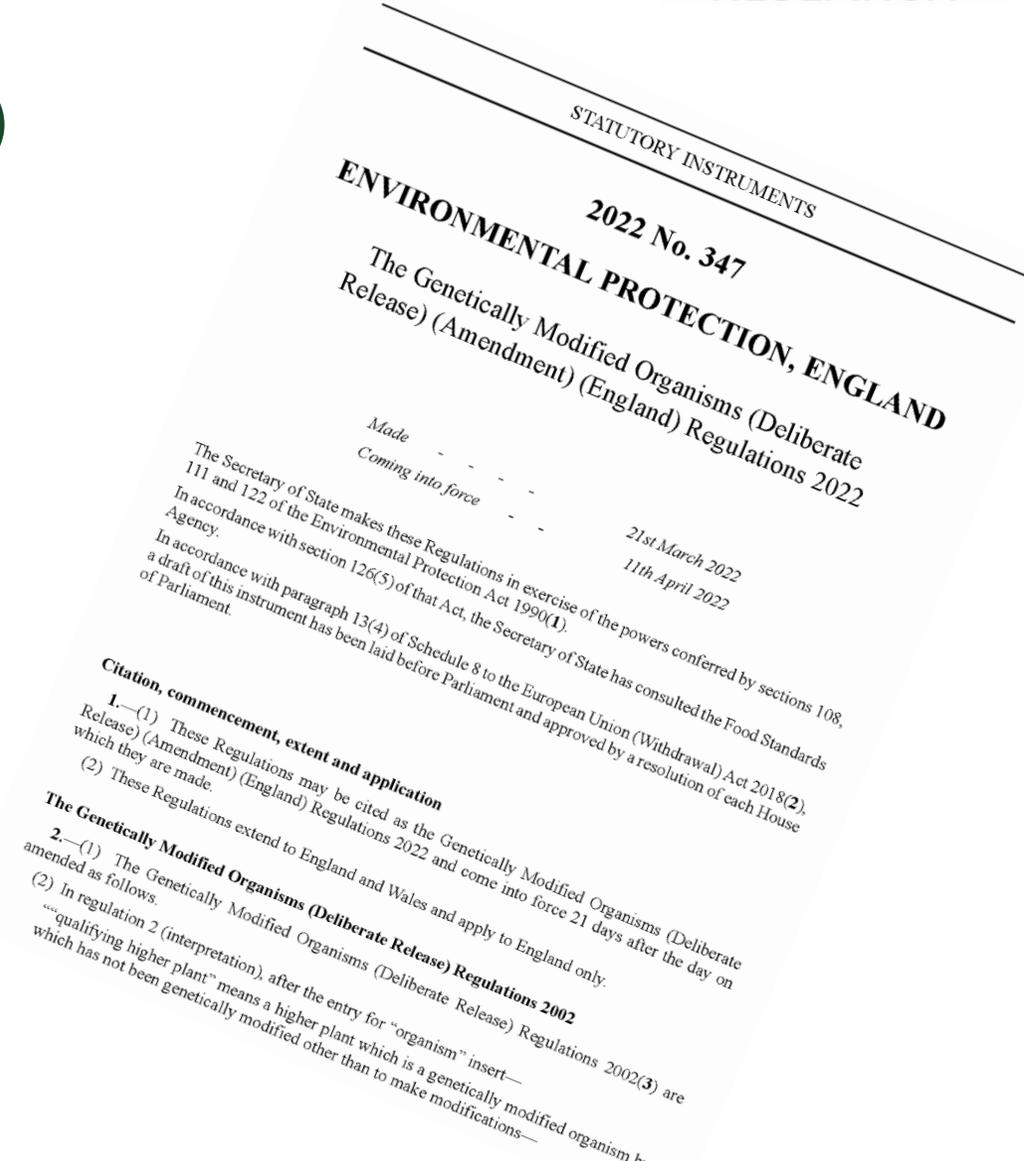
The Genetically Modified Organisms (Deliberate Release) Regulations 2002(3) are amended as follows.

(2) In regulation 2 (interpretation), after the entry for “organism” insert—

qualifying higher plant means a higher plant which is a genetically modified organism but which has not been genetically modified other than to make modifications:

(a) that could have occurred naturally, or

(b) that could have been made using one or more of the techniques set out in regulation 5(2);” [these include polyploidy induction, mutagenesis and cell/protoplast fusion].



Concluding remarks



ROTHAMSTED
RESEARCH



- Crop management best practice and the use of low asparagine wheat varieties are important aspects of regulatory compliance with respect to the acrylamide content of wheat products.
- Further step reductions in the free asparagine concentration of wheat grain are possible using genome editing and presumably also chemical mutagenesis.
- Effects on yield, N content, grain weight, grain protein etc. have still to be determined.
- This is important because genetic and agronomic approaches to solving the acrylamide problem could eventually lead to massive savings for food businesses, enabling them to comply with evolving regulations on acrylamide without costly changes to production lines or reduction in product quality.

Acknowledgements: Current low acrylamide wheat team



ROTHAMSTED
RESEARCH

Nigel Halford



Joe Oddy



Sarah Raffan



Acknowledgements: Academic and industry partners



ROTHAMSTED
RESEARCH

University of Reading

Steve Elmore

John Innes Centre

Simon Griffiths

Luzie Wingen

Shanghai Acad. Agricultural Sci.

Jianhua Huang

Zhiwei Chen

Chenghong Liu

Colorado State Univ.

Stephen Pearce

University of Bristol

Keith Edwards

Christy Waterfall

Paul Wilkinson

Gary Barker

Industry

AHDB, KWS UK Ltd., Saaten Union UK Ltd, RAGT
Seeds Ltd, Syngenta UK Ltd, Limagrain UK Ltd,
CEEREAL, Mondelez, Curtis Analytics, General Mills



Biotechnology and
Biological Sciences
Research Council